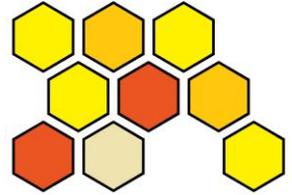


Great Hexpectations

(The RIBA Newsletter)



July 2019



Free bees! (photo: Emily Langlais)

Good summer tidings to you and your bees! This issue of the newsletter is coming to you a couple months later than I intended, but hopefully I can be excused for preferring the reality of working in the garden and among the bees over the act of writing about it. After all the miserable wet weather we had in the early spring, I am relishing sunny skies and blooming flowers when I finally have the leisure time to enjoy them. The picture above has been an unfortunately common sight for many over the last month. I was disappointed to see my champion honey-producing hive of 2018 swarm high into a tree at the end of May. We watched helplessly as the vortex of bees poured out, filling the air with a spectacular commotion of activity. As I write this on the first of July, we just finished up an inspection that turned up several capped swarm cells, and the queen produced by the earlier swarm, in the very same hive. Hopefully this time our manipulations will prove to be more successful!

The July newsletter features a thought-provoking article on bee education by Ed Syzmanski. Is a paradigm shift needed? It's a question that has been on my own mind often lately. Ed has also graciously agreed to ghostwrite this episode of the Beekeeper's Almanac. Jon Foster offers an overview of the honey bee dissection/microscopy class hosted by MDAR at UMass Amherst in April. Dr Jane Dennison reports on the completion of "the queen grant." Cindy Holt has an update on the hives at the RIBA Apiary in Johnston. Meeting recaps and upcoming events round out this issue.

-Scott Langlais, RIBA VP

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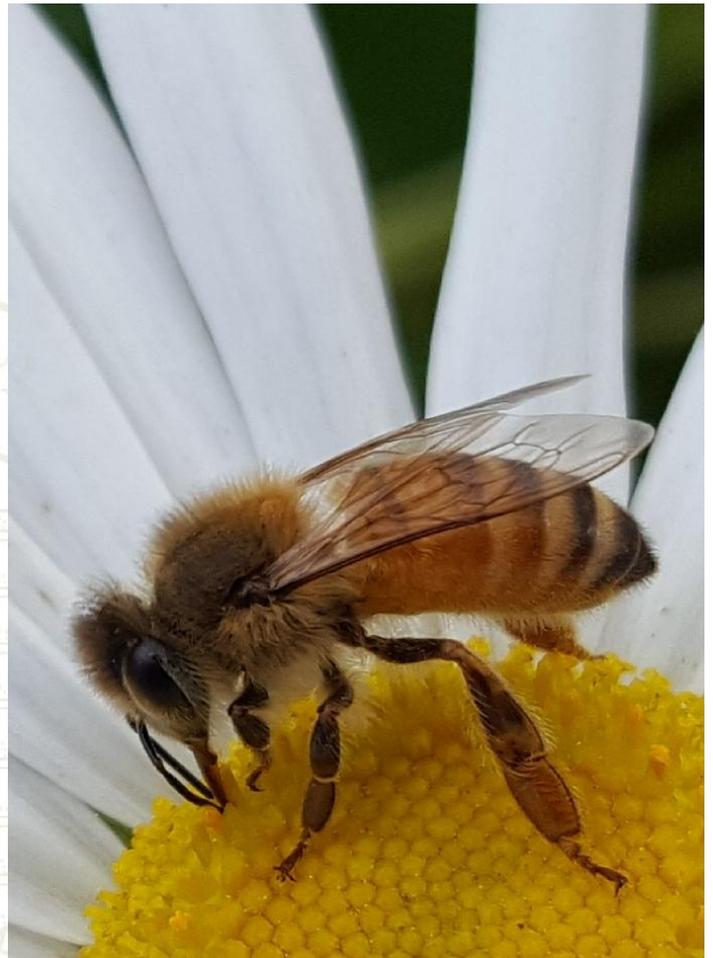
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Contributors to This Issue

Ed Szymanski, Cindy Holt, Jon Foster, Dr Jane Dennison, Emily Langlais, Scott Langlais

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-Scott Langlais

The RIBA LIBRARY

80+ Titles and growing!

RIBA is constantly adding to its collection of beekeeping literature, comprised of the most up-to-date works on a wide variety of subjects. These books are free to borrow for any valid RI library card-holder.

The books you want can be picked up from whatever branch is most convenient for you. Simply navigate to <https://catalog.oslri.net/> to begin your search. Enter keyword “RIBA” and narrow the focus to Greenville.

Accompanying photos are by the respective authors, except where noted.

Beekeeper's Almanac

Well, spring 2019 is behind us, and what a roller coaster it was! We went back and forth from wet to sunny to wet again, nectar was flowing, overwintered colonies grew rapidly, and then they got stuck inside for days at a time. That all adds up to the perfect recipe for swarming. And swarm they did – the first swarm reports came in around May 8th, and they just kept coming. Many of the swarms were retrieved - RIBA has some very resourceful beekeepers!

The very wet early spring posed particular challenges for new beekeepers installing packages. With little natural forage available, queens were slow to start laying, syrup-fed bees were building comb and filling it with syrup. When the sun finally came out and foragers starting bringing pollen in, queens had no place to lay eggs and the package bees were swarming! That, combined with other package queen issues, made it a rough start for the newbees this year.



For the overwintered colonies, the rain/sun/rain sequence meant copious nectar flowing, rapid development, and once the bees could spend some time out foraging, they really brought in the nectar. Many people are reporting early honey harvests, a welcome change from the past few years. I'm writing this on June 26 in Southern Massachusetts. The maples, fruit trees, early berries, and black locusts are finished. In my yard, we have several very heavy supers full of honey waiting to be fully capped. We'll be extracting some in a day or two. I walked around the yard this evening and made some flower observations. At the moment, we have blackberries, milkweed, thyme, roses, silver lace vine, and elderberries blooming. Our bees are visiting them, but the majority of foraging activity (and it's strong) right now is focused on the sumac and linden (basswood) trees in bloom nearby, adding to the nectar already being stored in the supers. It's important to have a diverse selection of food sources on hand for the bees – between the major nectar flows of the maples, black locusts, sumac, linden, etc, you can fill in with plantings in your yards. There are beekeepers anticipating a drying up of the incoming nectar when the lindens are finished, but I know from looking at my yard, there's still more to come. Walking around the yard tonight, I see a lot on the verge of blooming – winterberry, echinacea, hydrangea, mountain mint, veronica, late

raspberries, butterfly bush, porcelain berry. And more to come in the fall. As long as we don't have the late summer drought, we're in good shape for nectar and pollen availability for a while.

Many beekeepers have started doing mite counts, and so far the numbers have generally been low. Counts should be done monthly at a minimum. Consider sharing your test results with other beekeepers in your area. Don't hesitate to reach out for assistance if you need it.

Results of the latest Bee Informed Partnership survey indicate respondents experienced their worst overwintering success since the survey began in 2006. “Beekeepers across the United States lost 40.7% of their honey bee colonies from April 2018 to April 2019, according to preliminary results of the latest annual nationwide survey conducted by the University of Maryland-led nonprofit [Bee Informed Partnership](#). The survey results indicate winter losses of 37.7%, which is the highest winter loss reported since the survey began 13 years ago.” The announcement issued 6/19/19 by the University of Maryland can be read [here](#).

News like this is spurring discussions about making beekeeping more sustainable. The Norfolk County (MA) Beekeepers Association is holding meetings to discuss changing the club’s education philosophy to emphasize sustainable beekeeping practices.

More on that to come – in the meantime – happy summer, be sure to test for mites and treat when needed. Here’s hoping for a reasonable balance of sun and rain, and a continued flow of nectar.

-Ed Syzmanski



Left: viburnum, Right: sumac

Meeting Recaps

4/28/19, Special Meeting: Dr Jamie Ellis “Keeping Bees Alive”

RIBA was privileged enough to host Dr Jamie Ellis for two consecutive days in April. This special event was a bit of an experiment; we weren’t sure if the membership would come out for the same speaker two days in a row, or what the response to a Saturday meeting would be. Thankfully, both days were well attended, and the crowds were treated to one of the very best speakers in America.

Dr Ellis is the head of University of Florida’s Department of Entomology, the largest entomology department in the country. He is also an ordained Baptist minister, which likely explains part of why he is such an entertaining and engaging speaker. Saturday’s talk was an overview of several aspects affecting honey bee health in America. He began by pointing out the difference between gross vs. net losses in bee colonies (with the US experiencing an average 40% gross yearly loss, but a 1.1% net *gain*, due to beekeepers’ ability to split hives, catch swarms, etc).

The principle enemies of bee health are, he stated: 1) varroa and varroa-vectored pathogens, 2) poor nutrition, 3) queen problems, and (for commercial beekeepers) pesticides. “I am convinced that the average beekeeper doesn’t do enough to control varroa.” He recommended the Honey Bee Health Coalition’s [“Tools for Varroa Management,”](#) calling it the best document available for varroa control. His message was explicit: “failure to kill varroa means your bees are going to die.”

Most beekeepers need to be more aggressive at requeening failing or underperforming colonies. Newer beekeepers especially tend to develop attachments to queens, despite evidence of their lack of fitness. “We’re so happy to have a queen, we won’t fix the problem.” The keys to success in his view lay in understanding bee biology, and understanding your particular goals as a beekeeper. When those two principles are in harmony, we are on the right path.

4/29/19, Spring Banquet: Dr Jamie Ellis “Nucs: The Most Underutilized Tool in Beekeeping”

Our annual spring banquet was held at the Richmond Country Club in Richmond, RI. Dr Ellis’s talk was split into two separate sections. In the first half, he enumerated the many advantages of nucleus colonies. Briefly, these are:

1. Nucs alleviate swarming tendencies
2. Nucs can provide bees to help keep production colonies strong.
3. Nucs can provide a ready source of queens.
4. Nucs can be used as resources to aid weak colonies.
5. Nucs are an ideal way to make increase.
6. Selling nucs can be more profitable than selling honey.

In the second part of his time on stage, he discussed “The Biogeography of Honey Bees,” an entertaining look at the distribution of honey bee species around the world. Sources differ on how many species of honey bees exist globally, but Dr Ellis acknowledges nine: *Apis andreniformis*, *A. florea*, *A. dorsata*, *A. laboriosa*, *A. cerana*, *A. nuluensis*, *A. nigrocinta*, *A. koschevnikovi*, and *A. mellifera* (which is, of course, the one we generally concern ourselves with). Most of these species are native to Southeast Asia. Though there are currently no honey bees native to our continent, fossils of *Apis nearctica* have been found in North America, indicating that there were honey bees here at some point prior to the early American colonists’ importation of *Apis mellifera*. By his own admission, this talk was unlikely to aid beekeepers in their day-to-day management of their colonies, but the intent was to fire the imagination and share his obvious passion for these remarkable creatures.

5/19/2019: General Meeting

Master Beekeeper Sara Michaud opened the “newbies” meeting with a thorough overview of varroa, monitoring, and treatment techniques. Varroa can vector at least 8 viruses to honey bees, as well as PMS (parasitic mite syndrome). PMS affects the strongest colonies in late summer, since high populations of bees allow the highest populations of mites. These crashing colonies, left to their own fate, are the source of the “mite bombs” that can infect healthy neighboring colonies. An alcohol wash is considered the most reliable method of monitoring, per MA Department of Ag and ME Department of Ag. Visual checks, uncapping drone brood, and sticky boards are all considered



unreliable test methods. A 2-3% infestation rate is considered the threshold for treatment, depending on time of year. She admonished us not to rely on the touted hygienic behavior of specialty queens, saying “no bee is mite resistant” and “treatment free does not mean intervention free.” (see last page of the newsletter for Sarah’s chart of mite treatments)

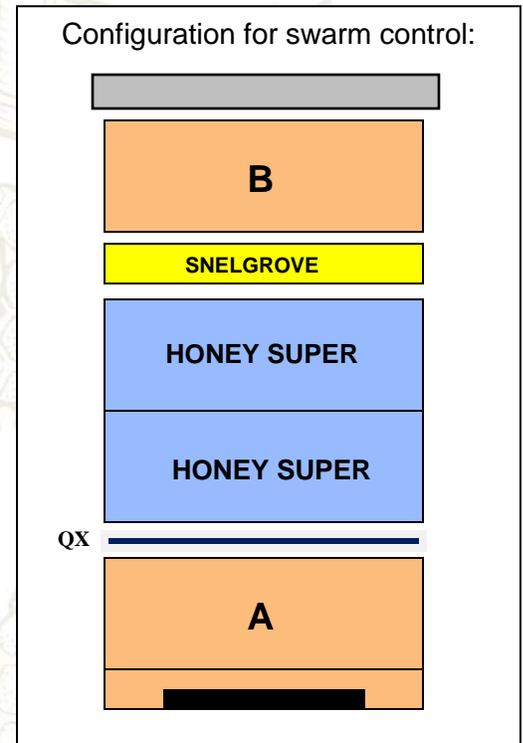
After the business portion of the meeting, our featured speaker was Roland Sevigny, a beekeeper with 47 years of experience and member of the Worcester County Beekeepers Association. His particular area of interest is the Snelgrove board aka double screen board. This piece of equipment has several interesting uses; among them are: swarm prevention, boosted honey production, and nuc/queen production.

Snelgrove board method of swarm control (see diagram):

Starting with a two-deep Langstroth, rearrange the boxes as follows: Box A goes on the original bottom board. The queen goes in box A, along with empty drawn comb, or foundation. Try to avoid putting any open brood in this box. Above box A is a queen excluder, then 2-3 honey supers. Above the honey supers we place the Snelgrove board, then box B. Box B should contain open brood, food, and especially frames with swarm cells. Open the right UPPER entrance on the Snelgrove board.

At this point box A should no longer wish to swarm; you’ve provided ample room for the queen to lay, removed swarm cells, and greatly reduced congestion in the hive. Foragers will continue to collect nectar and return to the original bottom board entrance, with plenty of space in the supers above the brood nest for incoming flow. Box B, separated by several boxes from the queen’s pheromones in box A, will likely perceive themselves to be queenless and should continue to care for the queen cell(s). If all goes as planned, box B will

raise their own mated queen in 3-4 weeks, depending on the age of the queen cell (you could also give box B a mated queen if you wanted to speed up the process; just be sure to cull any queen cells first).



As bees in the upper box (B) become foragers, they will be orienting to the right upper entrance. After several weeks this entrance will become crowded. This is where the Snelgrove board’s multiple entrances come into play. At this point, close the right upper entrance and open the right LOWER entrance. Returning foragers won’t realize the difference, but instead of entering box B, they will be channeled into the honey supers/box A. At the same time, we open the left UPPER entrance. New foragers from box B will recognize this as their entrance while older foragers will still be returning to the right side. By controlling which entrances in the Snelgrove board are open or shut we can selectively attenuate the strength of the foraging force in the top box, switching sides after 3-4 weeks.

This sounds confusing, but when you have the Snelgrove

board in your hands, you'll easily see how the upper/lower entrances can be manipulated to force the bees to enter either above or below it. It's certainly a bit more work (and thought) required than a simple walkaway split, but the benefit is that now you have two queens producing brood and contributing to the total foragers available to the hive, potentially leading to even greater total amounts of honey production.

Some further links:

<http://www.wbka.com/wp-content/uploads/2013/06/The-Many-Uses-Of-A-Snelgrove-Board-by-Wally-Shaw.pdf>

<https://honeybeesuite.com/how-to-over-winter-a-nuc/>

<https://honeybeesuite.com/uses-of-a-double-screen-board/>

http://pinkpages.chrisbacherconsulting.com/Imirie_Requeening_Method.html

(Editor's Note: The September 2018 issue of American Bee Journal also has an excellent article, with diagrams, on using the Snelgrove board for queen rearing.)

6/9/2019: 5th Annual RIBA Field Day, Salisbury Farm, Johnston, RI

I don't have a detailed recap of Field Day since I was one of the presenters and trying to keep one eye toward making sure the event ran smoothly. Instead, the following pages include a gallery of Emily's photos from the day. This year we tailored the presentations along three "tracks:" beginners, intermediate/advanced, and products of the hive. Each track featured three separate presentations. There were also three live bee demonstrations, our annual Smoker Contest (with proceeds going to benefit Special Olympics RI, hive tool sharpening, and a great day of fellowship among kindred spirits.



5th Annual RIBA Field Day, 6/9/2018



Photos: Emily Langlais

Towards More Sustainable Beekeeping

At the spring meeting of the Massachusetts Beekeepers Association, chief apiary inspector Kim Skyrn reported that it looks like losses for 2018-2019 in Massachusetts would exceed 50%, more like 55%. We've been running a steady 40-45% for many years and it seems to be getting worse. This is very upsetting to me - it's just not sustainable. Too many bees are dying. I think it's time for a change.

I'd like to see New England beekeeping associations take the lead in reducing our horrendous winter loss rates. In my own apiary, we have had years in the past when most or all of our bees died over the winter. It's devastating. We made some changes - we no longer buy bees with no mite resistant/tolerant or hygienic qualities, we buy local/resistant/tolerant/hygienic queens or raise our own, we treat for mites according to alcohol wash results. We read, talk to beekeepers in other clubs, and attend conferences and meetings. Our survival rate has improved dramatically over the past few years.

I think we need to adjust for changing conditions affecting our bees, from weather to nutrition to varroa mites. Lately, our spring seasons have been cold and wet; many of our bees are delivered in late March/early April. The weather is cool and damp, they develop slowly, can't get out to forage, and new beekeepers get frustrated with the process. Fall droughts are common; colonies are expected to overwinter on reduced stores, but some of our bees consume so much food that stores don't last through the winter. Varroa mites seem to be adapting to our treatment practices and multiply in the fall. Warmer fall weather, along with bees that raise brood into early winter, mean that varroa populations stay high later in the year, when treatment options are limited and winter bees' health can easily be compromised, as we have learned from Sam Ramsey.



What I'm suggesting is a change in philosophy. In Norfolk County Mass., our Varroa Control Group project broke new ground in beekeeper communication and coordinated treating. We're looking at expanding what has already been done into Rhode Island as a result of that. This program is a great start. The next step, in my opinion, is establishing coordinated and consistent education programs aimed at making beekeeping in New England more sustainable. We should take the opportunity to

make real change in the way we teach beekeepers, from the start right through more advanced education.

This isn't just a pipe dream of mine. If you talk to some of the more experienced and successful beekeepers, you'll find that many are on the same page. If you look at the SARE-funded research done by Erin Forbes on requeening southern packages and the many articles written by Randy Oliver, you find the same thread. Bees with no mite resistance or tolerance, no hygienic qualities, large food-consuming winter clusters, etc., all add up to a situation that is not sustainable in our climate, and it's getting worse with our changing climate. Many of our members buy nucs from local suppliers, but in populated areas the drone pools are still loaded with non-resistant bees and the good genetics are soon diluted. With smaller-scale communication groups and cooperation between beekeepers, the VCG program may allow us to work on controlling local drone populations for mite/virus resistance and genetic diversity.

...varroa populations stay high later in the year, when treatment options are limited and winter bees' health can easily be compromised...

Some argue that there aren't enough local bees to go around. I think we can find more local nuc suppliers and make those sources known to the clubs. Southern packages can be requeened with mite-tolerant queens. Packages are available from Northern California with Carniolan or Saskatraz queens. Others argue that higher quality bees are more expensive and new beekeepers spend enough money already. With increased success in overwintering, costs will decrease over time with fewer bees being purchased. And by raising our own queens and making our own nucs, a long term goal could be to never buy bees again, except maybe to buy a queen or two to add some new genetics.



...by raising our own queens and making our own nucs, a long term goal could be to never buy bees again...

This philosophy would begin with beginner Bee School and carry through our more advanced education. Newbees would be encouraged to buy local nucs to start, or requeen packages, and mentors can help with management. Follow-up classes for graduates can cover more advanced management. We can emphasize in our intermediate programs the use of tools like the Snelgrove board for avoiding swarms and raising queens, and using splits and raising queens for apiary expansion instead of just buying more bees.

We can continue the mite control education, have demonstrations through the VCG program, and have classes on philosophies and practices of sustainable beekeeping.

I hope to start some discussion about this in RIBA. You can email me at lunariafarm@outlook.com with comments.

-Ed Szymanski

Apis Mellifera Up Close.... Real Close!

When the University of Massachusetts offered its “Honey Bee Under the Microscope” class at its Amherst campus, I thought back to my high school biology class. The last time I was in a similar environment was some 45 years ago. During my winter bee reading of RIBA library books, I had stumbled on “Anatomy and Dissection of the Honeybee” by D. A. Dade. I quickly warmed to the idea of getting behind a microscope again. So on Saturday, April 6th, I joined with over 20 other enthusiastic “students,” including RIBA members Calvin Alexander, Keith Salisbury, Master Beekeepers Dane Pursley and Scott Langlais, in addition to five state bee inspectors from Massachusetts and Connecticut.

The class began promptly with an introduction to honey bee biology by a team of seasoned campus instructors who included veterinarian and beekeeper Professor Jean Mukherjee, from the infectious disease lab at UMass, Honey Bee Extension Educator Hannah Whitehead and Dr. Kim Skyrn, Chief Apiary Inspector for the Massachusetts Dept. of Agriculture.

The biology segment continued with a review of the use of the stereo (dissection) microscope, where specimens were viewed for the first time, up close--real close!

The actual dissection began by breaking down the external structures of a worker bee, removing the wings, legs, and separation of the head, thorax, and abdomen. We were all struck by all the hair that covers the bee, including the eyes, and marveled at the genetic engineering of this amazing



pollen and nectar gatherer. Once the head was removed, the details of the mandible and proboscis could be explored, as well as the trachea, between the head and thorax. It could determine by shape and color if that organ had been impacted by tracheal mites.

After a short lunch break, we were ready for a full afternoon of exploration.

It was my surprise (and relief) that the afternoon session of the class proved to be a “live” version of the D. A. Dade book. Many of the class handouts included the illustrations that were found in what I now fully understand to be a classic work.

Our real work for the afternoon focused on opening the abdomen to expose those internal organs, which included the digestive system, from the crop to the anus.

We were instructed in how to carefully use dissection tools to remove the series of plates (sclerites) interconnected by membranes. We viewed pollen being digested and the all important fat bodies, on which the varroa mite feeds. At the end of class, we prepared samples to be tested for nosema at the Beltsville Agricultural Research Center in Maryland. A compound microscope was set up so samples could be viewed before being sent out for evaluation.

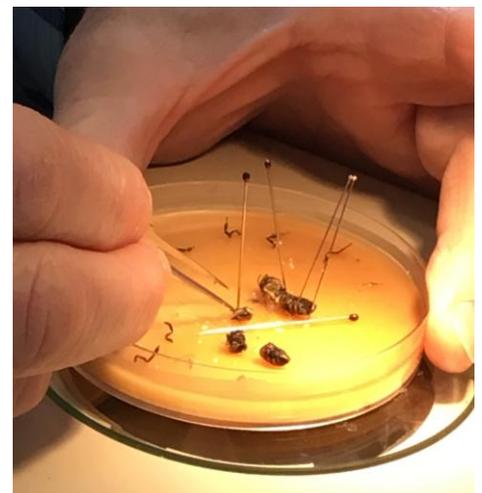
Calvin scored the first class “coup” under the microscope when his bee for dissection had a varroa mite still attached. For him, “the class brought full circle everything inside the bee and seeing what we are reading about and taught.”



Dane Pursley’s expectations of the class were completely satisfied. “I expected to learn basic dissection skills and the opportunity to view the honey bee’s internal organs under magnification. I was surprised by the vibrant detail visible under the microscope.”

Scott had mentioned the idea of our club acquiring microscopes, so I knew this class was a natural draw for his inquisitive mind. “I had seen some slides of honey bee internal organs under microscopy, but I was looking for a more in-depth experience. One of my long term goals for RIBA is to get a microscope that will be accessible to members. I thought this class would be an ideal way to pick up some information on the type of microscope that would be most useful to beekeepers, as well as some techniques that I could hopefully share with the membership.”

Dane summed up best the collective thoughts of our Amherst experience as “the instructors for the workshop presented a well planned and excellent program. Their willingness to share their expertise and knowledge helped me to increase my knowledge



which I, in turn, hope to pay forward to others.”

UMass-Amherst will be holding another “Honey Bee Under the Microscope” class in November 2019. For more information, contact: Hannah Whitehead, hwhitehead@umass.edu

-Jon Foster

Finishing Up the Pollinator Improvement Project (aka The Queen Grant)



-Emily Langlais

Happy summer to all!! As the first honey flow of the locust trees is underway, we need you all (who got a queen last year) to get back to us about the survival of those queens. This information will complete the grant requirements.

Spring 2018 queens came from two sources—100 New World Carniolans from HoneyRun Apiary and 100 Saskatraz from Cedar Lane Apiary.

Please contact Jane Dennison with the info about survival into the fall of 2018, and winter survival into early spring 2019. If you know the tag numbers (the metal tags on the hive and recorded in your apiary notes, give us that number also, along with your name and address. Reports can be sent to jamdmd@aol.com or texted to 401-374-9105.

Over the study years mated queens were obtained from the following queen producers:

- Cedar Lane Apiary (Saskatraz stock- saskatraz.com)
- Honeyrunapiaries.com (New World Carniolans—Sue Cobey stock)
- Lamb’s Honey Farm North Dakota, VPQueenbees.com (Italians with Pol-2.2 hygienic behavior)
- Wootensqueensbees.com (Italians from Northern California (Jackie Park-Burriss queen stock)
- B&B Honey Farms (Minnesota Hygienic stock from Marla Spivak, University of Minnesota)

These queens are available for private purchase if you want to continue to requeen with superior northern stock genetics. Lots of 25 are often less expensive, so team up with your friends!

The Pollinator Improvement Project brought 600+ mated queens with varroa hygienic behaviors into Rhode Island to increase the diversity of the drones that mate with swarm queens. Hopefully over the past two seasons, beekeepers who have participated in the grant became comfortable with the ability to make a split or nuc, requeen a hive,



-Emily Langlais

requeen a package, assess, treat and manage varroa. Please keep up the spirit of the grant. Requeen your southern stock hives with northern survivor queens, monitor and treat for mites, learn to make nucs to expand your colonies. Maybe someday soon the students coming out of bee school will have the ability to buy locally raised nucs and we can decrease the importation of southern packages with southern queens.

As the grant is now ended (once we get the survival data), you can move the hives to new locations, use drone trapping for varroa control (but that will decrease the drone genetics for wild mating), make nucs for sale---they are all your bees now!

Thank you all for your help and support during the grant seasons! Thanks to those who got the first year up and running in 2017, to the volunteers who handed out caged queens and did paperwork for registrations, the RIBA Board who kept my spirits up, Bob Barnes who is helping again gather the final data, Emily Patry who did all the scheduling for volunteers, Steve Burke who helped submit the 2018 results and hopefully will be there with his lawyerly and elegant prose as we give the final report, Ken Warchol the Mass Bee Inspector who ran educational sessions, DEM Snake Den and RIC Apiary for letting us use their real estate for queen banking and handouts, and most of all to Jim Lawson from RI DEM who went out of his way to help us maintain the queen banks and did off hours queen handouts, tracked and received valuable queen shipments from across the country, and explained over and over again the ways to help get the queens into their new RI hives. My very grateful thanks to ALL of you!

-Dr Jane Dennison



RIBA Apiary Update

Happy summer to you all!

I hope that that despite the cold wet spring, summer finds you with strong healthy colonies. This season has had its shares of ups and downs at the RIBA apiary, but all of the hives made it alive through the winter and into spring.

Patsy (we name our hives not our queens) was our most challenging hive, appearing queenless at first, only to discover after we added a new queen, that she actually had a drone laying queen. Her majesty was eventually located and now awaits her turn at dissection in our vice

president's freezer. We are happy to announce that the new queen, along with a nuc's worth of brood and bees, has been accepted and is getting down to business.

The Snelgrove board came in handy this spring as we took advantage of the swarming impulse to make several new queens to have on hand if needed. Through this we were able to requeen Patsy as well as Ivy. As of last week's inspection, we are happy to announce that all of our hives have laying queens and look to be in very good shape.

Dolly and Agnetha visited the RIBA field day this spring, to serve as demonstration hives. They enjoyed their little vacation at Salisbury farm and left before the bear that was sighted at the farm got a hold of them (phew).

The first round of Sentinel Apiary sampling was completed at the end of May. The results showed little to no mites and a slightly elevated level of nosema only in Dolly and Patsy. This has been the norm for the last several seasons in spring and we are expecting those levels to drop come next



Left: Cindy Holt, Calvin Alexander, Sue Wesley, Lizabeth Brown (Emily Langlais). Right: shaking bees at the entrance of the AZ hive (Cindy Holt)

round of results. Here's to another season of stuffing quarter cups of angry bees into jars with narrow necks! If you haven't already, be sure to look up the club hive at the Bee Informed Partnership Sentinel Apiary site. Our hive scale is not hard to find as we are the only one in Rhode Island.

The club's AZ hive is officially up and running at the yard! Thank you to Scott for doing such a spectacular job painting it! We were very fortunate to receive a ridiculously easy swarm call at the end of June. Emily and I were successful at wrangling the ladies and delivering them to the yard. I was very excited to stock the hive with a swarm, as they are best for drawing all the wax needed to get the hive ready for winter. Scott and I installed the swarm last week and I hope to see progress when we visit the yard later today. We hope to get some tips and tricks to share with you all in the future. In the meantime, happy beekeeping!

-Cindy Holt

Upcoming Events

- **July 15-19:** EAS, Greenville, SC <http://www.easternapiculture.org/conferences/eas-2019.html>
- **July 21:** General Membership Meeting, Snake Den State Park, Johnston RI
- **August:** *no RIBA meeting this month; too hot!*
- **September 8:** General Membership Meeting, home of Tom Chapman, West Greenwich, RI
- **October 27:** Fall Banquet, location TBD, speaker: Dr Keith Delaplane, University of Georgia (researcher and author of *First Lessons in Beekeeping*)
- **November 17:** General Membership Meeting, Guy Lefebvre Community Center, Coventry, RI; speaker: Larry Connor (author of many books/Wicwas Press)

Position to Fill

Eastern Apicultural Society Rhode Island Representative

- three year term
- board meetings at EAS conference and twice a year in person or via conference call.
- contribute RIBA activities to the meetings/conference pages for Bee Culture magazine.
- learn how EAS functions as the EAS summer conferences are planned.
- take a basket of goodies/ foodstuffs/gifts to represent RI for the fundraiser auction at the EAS Conference. (RIBA will reimburse \$)
- meet great people who make a difference to a dynamic organization
- talk up EAS conference to RIBA
- be willing to attend (at own expense) the summer conference
- submit the RI dues invoice for EAS to the RIBA secretary.

Contact: Dr Jane Dennison jamdmd@aol.com or text to 401-374-9105



-Emily Langlais

Mite Treatments

Chemical	Active Ingredient	Temperature	Method	Treatment duration	Treat with supers on?	Time to wait for supers	Max. applications per year	Extra Precautions
Apiguard	Thymol	60-105	Gel tray on brood frames	28 days	No	Immediately		Do not eat honey from brood nest
Api Life Var	Thymol, eucalyptus oil, menthol	65-85	Tablets in corners of brood frames	26-32 days	No	1 month	2x/yr	Combustable Don't apply if robbing evident
Mite Away Quick Strips	Formic acid	50-85 (above 92 degrees, increased queen loss)	Pads on brood nest	7 days (1/2 dose every 2-6 weeks)	Yes	Left on		Wait 3 days after inspection to apply, then do not open after application until removal
Oxalic acid	Oxalic acid dehydrate	Dribble: above 37 Vapor: N/A	Dribble brood nest, vaporize entrance	10 minutes	No	2 weeks	2x/yr	No brood (not registered in US to use with brood) Do not eat honey from brood nest
Hop Guard II	Hops Beta acids	N?A	Strips in brood nest (NO strips in supers)	30 days	Yes	Left on	3x/yr	Do not eat honey from brood box, Can treat packages with different dosing.
Apivar	Amitraz	N/A Not in winter	Strips in brood nest	42-56 days	No	2 weeks	2x/yr	Do not eat honey from brood nest
Apistan	Tau-fluvalinate	Above 50	Strips in brood nest	42-56 days	No	Immediately	2x/yr	Do not consume wax or honey from brood nest

-Sara Michaud

